Oytodelics

Whole blood processing kit

P03 - Fresh blood fixation

- 1. **Calculate** the required volume of the **Fixation Buffer** (total volume of blood processed within one batch multiplied by factor 10) and **prepare** it by mixing equal volumes of
 - a. Fix-concentrate
 - b. Fix-Diluent
- 2. **Dispense** the **Fixation Buffer** (10-times volume of preserved blood) to tubes and let it equilibrate to room temperature (approx. 5 min).
- 3. **Transfer** blood samples to the tube with the Fixation buffer, set the timer, and **incubate** for **15 minutes** at **room temperature**. Vortex multiple times throughout.
- 4. **Add** 1-times concentrated **LYSIS buffer** (40-times volume of the blood sample ¹), **mix** well by inverting tube multiple times, and **incubate** at room temperature for minimum **15 to 20 minutes** until a complete erythrocytes' lysis.
- 5. **Centrifuge** at **300g for 5 to 10 minutes**².
- 6. **Aspirate the supernatant** down to:
 - a. "ring" on the 5mL "FACS" tube leaving minimum 100 μ l of supernatant over the pellet,
 - b. approx. mark 0.5 ml in 15mL tube (for samples with original volume up to 250µl blood processed in the 15mL tube from beginning),
 - c. approx. mark 5.0-7.5 ml in 50mL tube, and before the next step, proceed as follows:
 - i. transfer to a 15mL tube
 - ii. centrifuge at 300g for 5 minutes,
 - iii. aspirate the remaining volume down to approx. mark 0.5 ml.
- 7. Break the pellet by vortexing and **add** 1-time concentrated **WASH buffer** (40 times the volume of the blood sample³).

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¹ E.g., for 1.0 ml of blood and 10.0 ml of Fixation Buffer use **40.0 ml of the LYSIS buffer**.

 $^{^2}$ Set the centrifugation timing according to the liquid column height in the tube and rotor size. The timing should increase with shorter rotor diameter and/or more liquid (higher liquid column) per tube.

³ E.g., for 1.0 ml of blood, 10.0 ml of Fixation Buffer, and 40ml of LYSIS buffer use **40.0 ml of the WASH buffer**.

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- 8. Centrifuge at 300g for 5 to 10 minutes4.
- 9. **Gently aspirate** the supernatant down to:
 - a. approx. mark 0.25 ml (15mL tube)
 - b. ring" on the 5mL "FACS" tube leaving approx. 100 μ l of supernatant over the pellet.

Do not decant!

- 10. Proceed with:
 - a. acquisition
 - b. **cryogenic preservation** in CRYO#20 buffer at ≤ -80°C. For thawing, proceed as follows:
 - i. Thaw the sample in a range from 20°C to 37°C (ideally on a heat block),
 - ii. Acquire ⁵ samples or continue with staining procedure without washing ⁶ step.



⁴ Set the centrifugation timing according to the liquid column height in the tube and rotor size. The timing should increase with shorter rotor diameter and/or more liquid (higher liquid column) per tube.

⁵ Keep the volume of the CRYO#20 buffer unchanged or optimise it according to your flow cytometer specifications.

 $^{^6}$ Centrifugate the sample at 300-400g/ \approx 5 min and remove some of the supernatant to optimize volume for staining with Ab-cocktail.

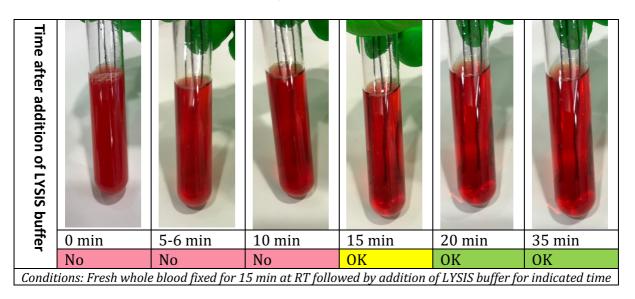
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Scaling ratios							
Blood	1	:	10	Fixation buffer			
Blood	1	:	40	LYSIS buffer			
Blood	1	:	40	WASH buffer			
Blood	1	:	0.5 - 1	CRYO#20			

Examples of recommended processing conditions										
Blood	Fix LYSIS		WASH	Recommended processing tube size		CRYO#20				
sample	buffer	buffer	buffer	First wash	All consequent	volume				
volume	volume	volume	volume		washes					
100 μl	1.0 ml	4.0 ml	4.0 ml	10 – 15 ml	5-15 ml	100 μl				
250 μl	2.5 ml	10 ml	10 ml	15 ml	15 ml	100 μl				
500 μl	5.0 ml	20.0 ml	20.0 ml †	50 ml *	15 ml	250 μl				
1.0 ml	10.0 ml	39.0 ml	40.0 ml †	50 ml *	15 ml	500 μl				

^{*} To achieve optimal cell yields execute only first spin after Fix&Lyse step in 50 ml tube, aspirate to mark 5 ml or above (not below 5 ml mark), transfer to 15 ml tube and continue with altered WASH buffer volumes.

Guide on decision of RBCs lysis step duration:



[†] In case you decide to run all consequent processing steps in 15 ml tube, use 14 ml of WASH buffer and adjust number of washing steps based on pellet color. Typically, only one more washing step is required.