P01 - Sample stabilisation

Preparation phase:

Aliquot cryogenic vials[1] with **Cytodelics Stabiliser equal the planned blood sample volume**. ***Store at 5–8˚C*** *until sampling (up to* ***2 years****). Identify dispensed volume on the label.* Use the Human Stabiliser (hWBCS) for human blood and the Murine Stabiliser (mWBCS) for mouse or rat blood.

Protocol:

1. Bring tubes pre-filled with the Stabiliser **to room temperature** (5 – 10 minutes).
2. Transfer equal **blood** volume to the cryogenic vial (RATIO 1:1).
3. **Mix** the blood sample with the Stabiliser **by flipping** the vial 10-15 times. **Do not** vortex!
4. Incubate at room temperature for 10 minutes.
5. Store the sample until further processing at:
    a. **≤ -80˚C** (long-term storage; whole blood stored for up to 2.5 years kept a stable immune profile in an independent study[1]);
    b. **≤ -20˚C** (short term, up to 3 weeks), then transfer to the ultra-low freezer (**≤ -80˚C**) as soon as possible and keep there until further processing;
    c. or place on **dry ice** and transfer to the ultra-low freezer (**≤ -80˚C**) as soon as possible and keep there until further processing.

[1] *Parthasarathy V. et al., Cytometry Part A (2026), doi: 10.1002/cyto.a.70059.*

[1] Choose cryogenic vial with nominal volume of approx. 3-times of the planned blood sample.

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P01 - Processing after sample thawing

1. **Calculate** the required volume of the **Fixation Buffer** (total volume of blood processed within one batch multiplied by factor 10) and **prepare** it by mixing equal volumes of
   a. Fix-concentrate
   b. Fix-Diluent
2. **Dispense** the **Fixation Buffer** (10-times volume of preserved blood; do not count on the Stabiliser volume[2]) to tubes and let to equilibrate to room temperature (approx. 5 minutes).
3. **Thaw** blood samples preserved in the Stabiliser by quickly heating on water bath or heat block set to 37℃ (approx. 1 minute).
4. **Transfer** {blood+Stabiliser} mixture to the tube with the Fixation buffer, set the timer, and **incubate** for **15 minutes** at **room temperature**. Vortex multiple times throughout.
5. **Add** 1-time concentrated **LYSIS buffer** (40-times volume of the blood sample[3]), **mix** well by inverting tube multiple times, and **incubate** at room temperature for minimum of **15 to 20 minutes** until a complete erythrocytes' lysis.
6. **Centrifuge** at **300g for 5 to 10 minutes**[4].
7. **Aspirate the supernatant,** according to the size of the sample, down to**:**
   - approx. mark 5.0-7.5 ml in 50mL tube, and before the next step, proceed as follows:
     i. transfer to a 15mL tube,
     ii. rinse twice with 2.0 ml of the **LYSIS buffer** and transfer to the 15mL tube,
     iii. centrifuge at 300g for 5 minutes,
     iv. aspirate the remaining volume down to approx. mark 0.5 ml.
   - approx. mark 0.5 ml in a 15mL tube (for samples with original volume up to 250µl blood processed in the 15mL tube from the beginning),

[2] E.g., for 1.0 ml of blood preserved in 1.0 ml of Stabiliser use **10.0 ml of the Fixation Buffer**, i.e., 10-times volume of the blood.
[3] E.g., for 1.0 ml of blood and 10.0 ml of Fixation Buffer use **40.0 ml of the LYSIS buffer**.
[4] Set the centrifugation timing according to the liquid column height in the tube and rotor size. The timing should increase with shorter rotor diameter and/or more liquid (higher liquid column) per tube.

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- "ring" on the 5mL "FACS" tube, leaving a minimum of 100 µl of supernatant over the pellet.

8. Break the pellet by vortexing and **add** 1-time concentrated **WASH buffer** (40-times the volume of the blood sample[5]).
9. **Centrifuge** at **300g for 5 to 10 minutes**[6].
10. **Gently aspirate** the supernatant down to:
    a. approx. mark 0.25 ml (15mL tube)
    b. ring" on the 5mL "FACS" tube leaving approx. 100 µl of supernatant over the pellet.

    Do not decant!
11. Proceed with:
    a. sample staining and acquisition,
    b. **cryogenic preservation** in CRYO#20 buffer at ≤ –80°C.

    For thawing, proceed as follows:
    i. Thaw the sample in a range from 20°C to 37°C (ideally on a heat block),
    ii. Process according to your downstream application[7].

[5] E.g., for 1.0 ml of blood, 10.0 ml of Fixation Buffer, and 40ml of LYSIS buffer use **40.0 ml of the WASH buffer**.
[6] Set the centrifugation timing according to the liquid column height in the tube and rotor size. The timing should increase with shorter rotor diameter and/or more liquid (higher liquid column) per tube.
[7] You may stain without the washing step and acquire; however, we recommend that you test compatibility of your staining panel. Before the staining, centrifuge the sample at 300-400g/≈5 min and remove some of the supernatant to optimise volume of the Ab-cocktail and antibodies consumption.

Scaling ratios		
Blood	1 : 1	Cytodelics Stabiliser
Blood	1 : 10	Fixation buffer
Blood	1 : 40	LYSIS buffer
Blood	1 : 40	WASH buffer
Blood	1 : 0.5 - 1	CRYO#20

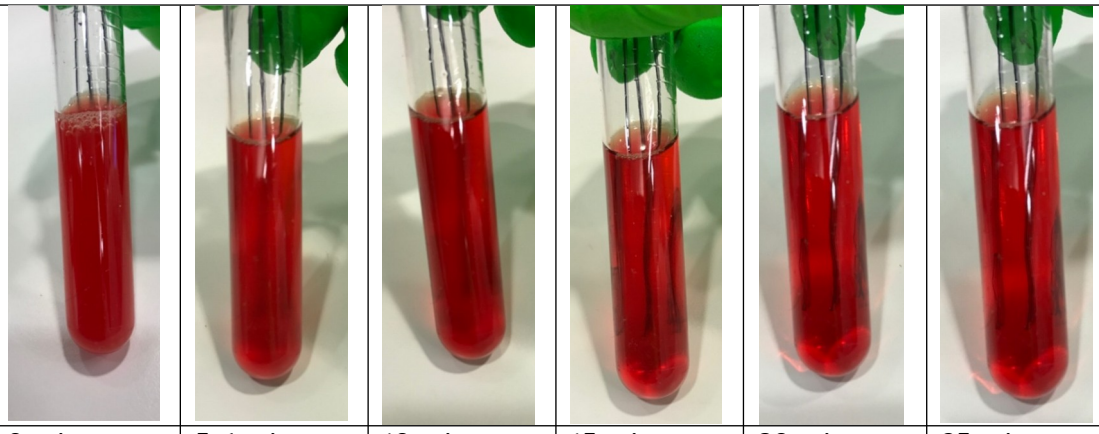
Examples of recommended processing conditions

Blood sample volume	**Fix** buffer volume	**LYSIS** buffer volume	**WASH** buffer volume	**Recommended processing tube size**		**CRYO#20** volume
				First wash	All consequent washes	
100 µl	1.0 ml	4.0 ml	4.0 ml	10 – 15 ml	5-15 ml	100 µl
250 µl	2.5 ml	10 ml	10 ml	15 ml	15 ml	100 µl
500 µl	5.0 ml	20.0 ml	20.0 ml †	50 ml *	15 ml	250 µl
1.0 ml	10.0 ml	39.0 ml	40.0 ml †	50 ml *	15 ml	500 µl

* To achieve optimal cell yields execute only first spin after Fix&Lyse step in 50 ml tube, aspirate to mark 5 ml or above (not below 5 ml mark), transfer to 15 ml tube and continue with altered WASH buffer volumes.

† In case you decide to run all consequent processing steps in 15 ml tube, use 14 ml of WASH buffer and adjust number of washing steps based on pellet color. Typically, only one more washing step is required.

Guide on decision of RBCs lysis step duration:



Conditions: Fresh whole blood fixed for 15 min at RT followed by addition of LYSIS buffer for indicated time